

## **Olerup SSP<sup>®</sup> DQB1\*02**

Product number:	101.213-24u – without <i>Taq</i> polymerase
Lot number:	77K
Expiry date:	2013-July-01
Number of tests:	24
Number of Wells per test:	6
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

**This Product Description is only valid for Lot No. 77K.**

### **CHANGES COMPARED TO THE PREVIOUS *OLERUP SSP<sup>®</sup>* DQB1\*02 Lot**

The DQB1\*02 specificity and interpretation tables have been updated with the DQB1 alleles described since the previous *Olerup SSP<sup>®</sup>* DQB1\*02 lot (**Lot No. 08G**) was made.

The DQB1\*02 primer set is unchanged compared to the previous lot.

## PRODUCT DESCRIPTION

### DQB1\*02 SSP subtyping

#### CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1\*02:01 to DQB1\*02:05 alleles.

*Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.*

#### PLATE LAYOUT

Each test consists of 6 PCR reactions in an 8 well PCR plate. Wells 7 to 8 are empty.

1	2	3	4	5	6	empty	empty
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The 8 well PCR plate is marked with 'DQ2' in silver/gray ink.

Well No. 1 is marked with the Lot No. '77K'.

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are heat-sealed with a PCR-compatible foil.

**Please note:** When removing each 8 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

#### INTERPRETATION

Only DQB1 alleles will be amplified by the DQB1\*02 subtyping kit. Thus, the interpretation of DQB1\*02 subtypings is not influenced by the DQB2 and DQB3 genes.

#### UNIQUELY IDENTIFIED ALLELES

All the DQB1\*02 alleles, i.e. **DQB1\*02:01 to DQB1\*02:05**, recognized by the HLA Nomenclature Committee in October 2010<sup>1</sup> will give rise to unique amplification patterns by the primers in the DQB1\*02 subtyping kit.

The DQB1\*02 kit cannot distinguish the DQB1\*02:01-02:01:03 alleles.

<sup>1</sup>HLA-DQB1 alleles listed on the IMGT/HLA web page 2010-October-15, release 3.2.0, [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla).

#### RESOLUTION IN HOMO- AND HETEROZYGOTES

A total of 7 alleles generate 5 amplification patterns that can be combined in 15 homozygous and heterozygous combinations. 4 of these genotypes do not give rise to unique amplification patterns.

+++++                    \*02:01:01, \*02:04 = \*02:04, \*02:04

+++++                    \*02:01:01, \*02:05 = \*02:05, \*02:05

\*02:01:01 = \*02:01:01-02:01:03

## SPECIFICITY TABLE

### DQB1\*02 SSP subtyping

Specificities and sizes of the PCR products of the 6 primer mixes used for DQB1\*02 SSP subtyping

Primer Mix	Size of spec. PCR product <sup>1</sup>	Size of control band <sup>2</sup>	Amplified DQB1*02 alleles	Other amplified DQB1 alleles <sup>3</sup>
1	120 bp	515 bp	*02:01:01-02:02, 02:04-02:05	
2 <sup>4</sup>	85 bp	430 bp	*02:03	
3	145 bp	515 bp	*02:01:01- 02:01:03, 02:04- 02:05	*03:01:01-03:23, 03:25-03:31, 04:01:01-04:06, 05:01:01-05:08, 06:01:01-06:37, 06:39-06:40
4	140 bp	430 bp	*02:02-02:03	
5	145 bp	430 bp	*02:04	
6	180 bp	430 bp	*02:05	

<sup>1</sup>Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQB1\*02 SSP typings.

When the primers in a primer mix can give rise to specific PCR products of more than one length this is indicated if the size difference is 20 base pairs or more. Size differences shorter than 20 base pairs are not given. For high resolution SSP kits the respective lengths of the specific PCR product(s) of the alleles amplified by these primer mixes are given.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

<sup>2</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1\*02 subtyping.

In addition, well number 3 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

In the presence of a specific amplification the intensity of the control band often decreases.

<sup>3</sup>For several DQB1 alleles only partial 3<sup>rd</sup> exon nucleotide sequences are available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of the 3<sup>rd</sup> exon of DQB1 alleles are conserved within allelic groups.

<sup>4</sup>Short specific PCR fragments are less intense and not as sharp as longer specific bands.

<b>INTERPRETATION TABLE</b>						
<b>DQB1*02 PCR-SSP subtyping</b>						
<b>Allele-specific amplification patterns of the 02:01 to 02:05 alleles</b>						
	<b>Well</b>					
	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>
<b>Length of spec.</b>	<b>120</b>	<b>85</b>	<b>145</b>	<b>140</b>	<b>145</b>	<b>180</b>
<b>PCR product</b>						
<b>Length of int.</b>	<b>515</b>	<b>430</b>	<b>515</b>	<b>430</b>	<b>430</b>	<b>430</b>
<b>pos. control<sup>1</sup></b>						
<b>5'-primer<sup>2</sup></b>	<b>30(185)</b>	<b>57(266)</b>	<b>135(500)</b>	<b>102(400)</b>	<b>136(502)</b>	<b>30(185)</b>
	5' -AAg 3'	5' -TgA 3'	5' -TgA 3'	5' -TCT 3'	5' -ACg 3'	5' -AAg 3'
<b>3'-primer<sup>3</sup></b>	<b>57(266)</b>	<b>71(309)</b>	<b>170(604)</b>	<b>135(500)</b>	<b>170(604)</b>	<b>77(326)</b>
	5' -Cgg 3'	5' -CgT 3'	5' -gAC 3'	5' -ggC 3'	5' -gAC 3'	5' -CCg 3'
<b>Well No.</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>
<b>DQB1 allele<sup>4</sup></b>						
<b>*02:01:01-02:01:03</b>	<b>1</b>		<b>3</b>			
<b>*02:02</b>	<b>1</b>			<b>4</b>		
<b>*02:03</b>		<b>2</b>		<b>4</b>		
<b>*02:04</b>	<b>1</b>		<b>3</b>		<b>5</b>	
<b>*02:05</b>	<b>1</b>		<b>3</b>			<b>6</b>
<b>*03:01:01-03:23, 03:25-03:31, 04:01:01-04:06, 05:01:01-05:08, 06:01:01-06:37, 06:39-06:40</b>			<b>3</b>			
<b>DQB1 allele<sup>4</sup></b>						
<b>Well No.</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>

<sup>1</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1\*02 subtyping.

In addition, well number 3 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

<sup>2</sup>The codon, and in parenthesis the nucleotide, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon, matching the specificity-determining 3'-end of the primer is given. Codon and nucleotide numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>3</sup>The codon, and in parenthesis the nucleotide, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Codon and nucleotide numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>4</sup>For several DQB1 alleles only partial 3<sup>rd</sup> exon nucleotide sequences are available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of the 3<sup>rd</sup> exon of DQB1 alleles are conserved within allelic groups.

CELL LINE VALIDATION SHEET										
DQB1*02 SSP subtyping kit										
				Production No.	Well					
					1	2	3	4	5	6
					200961701	200961702	200961703	200961704	200961705	200961706
	IHWC cell line		DQB1							
1	9001 SA		*05:01		-	-	+	-	-	-
2	9280 LK707		*06:01	*02:02	+	-	+	+	-	-
3	9011 E4181324		*06:01		-	-	+	-	-	-
4	9275 GU373		*02:01		+	-	+	-	-	-
5	9009 KAS011		*05:02		-	-	+	-	-	-
6	9353 SM		*03:02	*06:01	-	-	+	-	-	-
7	9020 QBL		*02:01		+	-	+	-	-	-
8	9025 DEU		*03:01		-	-	+	-	-	-
9	9026 YAR		*03:02		-	-	+	-	-	-
10	9107 LKT3		*04:01		-	-	+	-	-	-
11	9051 PITOUT		*02:02		+	-	-	+	-	-
12	9052 DBB		*03:03		-	-	+	-	-	-
13	9004 JESTHOM		*05:01		-	-	+	-	-	-
14	9071 OLGA		*04:02		-	-	+	-	-	-
15	9075 DKB		*03:03		-	-	+	-	-	-
16	9037 SWEIG007		*03:01		-	-	+	-	-	-
17	9282 CTM3953540		*02:01	*06:03	+	-	+	-	-	-
18	9257 32367		*06:02	*02:02	+	-	+	+	-	-
19	9038 BM16		*03:01		-	-	+	-	-	-
20	9059 SLE005		*06:04		-	-	+	-	-	-
21	9064 AMALA		*03:01		-	-	+	-	-	-
22	9056 KOSE		*05:03	*06:04	-	-	+	-	-	-
23	9124 IHL		*05:03	*06:01	-	-	+	-	-	-
24	9035 JBUSH		*03:01		-	-	+	-	-	-
25	9049 IBW9		*02:02		+	-	-	+	-	-
26	9285 WT49		*02:01		+	-	+	-	-	-
27	9191 CH1007		*04:01	*05:01	-	-	+	-	-	-
28	9320 BEL5GB		*02:02	*03:01	+	-	+	+	-	-
29	9050 MOU		*02:02		+	-	-	+	-	-
30	9021 RSH		*04:02		-	-	+	-	-	-
31	9019 DUCAF		*02:01		+	-	+	-	-	-
32	9297 HAG		*03:01		-	-	+	-	-	-
33	9098 MT14B		*03:02		-	-	+	-	-	-
34	9104 DHIF		*03:01		-	-	+	-	-	-
35	9302 SSTO		*03:05		-	-	+	-	-	-
36	9024 KT17		*03:02		-	-	+	-	-	-
37	9065 HHKB		*06:03		-	-	+	-	-	-
38	9099 LZL		*03:01		-	-	+	-	-	-
39	9315 CML		*02:01	*03:01	+	-	+	-	-	-
40	9134 WHONP199		*02:02	*03:03	+	-	+	+	-	-
41	9055 H0301		*06:09		-	-	+	-	-	-
42	9066 TAB089		*06:01		-	-	+	-	-	-
43	9076 T7526		*03:03		-	-	+	-	-	-
44	9057 TEM		*05:03		-	-	+	-	-	-
45	9239 SHJO		*02:02		+	-	-	+	-	-
46	9013 SCHU		*06:02		-	-	+	-	-	-
47	9045 TUBO		*03:01		-	-	+	-	-	-
48	9303 TER-ND		*05:01		-	-	+	-	-	-

## CERTIFICATE OF ANALYSIS

### **Olerup SSP® DQB1\*02 SSP**

**Product number:** 101.213-24u– without *Taq* polymerase  
**Lot number:** 77K  
**Expiry date:** 2013-July-01  
**Number of tests:** 24  
**Number of wells per test:** 6

#### **Well specifications:**

Well No.	Production No.
1	2009-617-01
2	2009-617-02
3	2009-617-03
4	2009-617-04
5	2009-617-05
6	2009-617-06

The specificity of each primer solution of the kit has been tested against 48 well characterized IHWC cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 2, 5 and 6 were available. The specificities of the primers in primer solutions 2 and 6 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer. In primer solution 5 it was only possible to test the 3'-primer, the 5'-primer was not possible to test.

**Results:** No false positive or false negative amplifications were obtained.

**Date of approval:** 2011-January-14

**Approved by:**

**Quality Control, Supervisor**

## Declaration of Conformity

**Product name:** *Olerup* SSP® DQB1\*02  
**Product number:** 101.213-24u  
**Lot number:** 77K

**Intended use:** DQB1\*02 high resolution histocompatibility testing

**Manufacturer:** *Olerup* SSP AB  
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We, *Olerup* SSP AB, hereby declare that this product, to which this Declaration of Conformity relates is in conformity with the following Standard(s) and other normative document(s) ISO 9001:2008 and ISO 13485:2003, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices, Annex III, as transposed into the national laws of the Member States of the European Union.

The Technical Documentation File is maintained at *Olerup* SSP AB, Hasselstigen 1, SE-133 33 Saltsjöbaden, Sweden.

The Authorized Representative located within the Community is: *Olerup* SSP AB.

Saltsjöbaden, Sweden  
2011-January-14

Olle Olerup  
Managing Director

Lot No.: **77K**

Lot-specific information

[www.olerup-ssp.com](http://www.olerup-ssp.com)

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